

3. Xu H, Chan Y, Bradel-Tretheway B, Akvol-Ataman Z, Skiniotis G, Lee B, Zhou ZH, Broder CC. A Novel Hexamer of Innate Immunity Nipah Virus Fusion Glycoprotein Reveals a Novel Hexamer of Innate Immunity. *J Virol* 2016; 80(12):e1005322. doi:10.1128/JVI.01331-16. PMID: 26919959
4. Mire CE, Satterfield BA, Geisbert JB, Agans KN, Borisevich M, Chen YD, Gao GDS, Fritch KA, Broder CC, Geisbert TW (2016). Pathogenic Differences between Nipah Virus Bealadeh and Malaysia Strains in Primates: Implications for Antibody Therapy. *Sci Rep* 3:6:30916. doi:10.1038/srep30916.

B. Positions and Honors

Positions and Honors

- 1990 -92 National Research Council of Canada, Associate, Laboratory of Allergy and Infectious Diseases, National Institute of Health, Bethesda, Maryland.
- 1992 -06 IRTA Fellow, V.D. NIAID, Krieger, Bethesda, Maryland.
Assistant Professor, Department of Microbiology and Immunology, Joint appointment, Molecular and Cell Biology Graduate Program, Uniformed Services University, Maryland.
- 2000 -05 Associate Professor, Department of Microbiology and Immunology, Joint appointment, Emerging Infectious Diseases Graduate Program, USUHS, Bethesda, Maryland.
- 2005 - Present Professor, Department of Microbiology and Immunology, Joint appointment, Emerging Infectious Diseases Graduate Program, USU, Bethesda, Maryland.
- 2006 -18 Director, Emerging Infectious Diseases Graduate Program, USU, Bethesda, Maryland.
- 2018 - Present Chair, Department of Microbiology and Immunology, USU, Bethesda, Maryland.

Other Experience and Professions

- 2009 Member, National Veterinary Stockpile Nipah Virus, Australia.
- 2011 Member, Discontools Nipah virus infection panel Expert Group, Gap analysis. International Federation for Animal Health Europe, Brussels, Belgium
- 2011 Invited expert, National Academies, Washington, DC. Evaluation of site-specific risk assessment for the National Bio- and Agro-Defense Facility (NBAF), Manhattan, Kansas

Editorial Board of *J. of Virology*

Honor and Awards

- 1996 The Henry W. Award for Excellence in Basic Science Research
- 1996 American Association for the Advancement of Science, Newcomb Cleveland Prize Magazine, Newcomb Cleveland Prize
- 1996 Outstanding Instructor in Virology, USUHS, School of Medicine
- 2005 The Henry W. Award for Excellence in Basic Science Research
- 2013 The Sidney Pestka Lecture, 22nd Annual Plenary Meeting, American Society for Virology
- 2013 The 2013 Federal Laboratory Consortium (FLC) Award for Excellence in Technology Transfer
- 2013 Second Finalist for the CSIRO Chair of Excellence in Technology and Innovation
- 2013 The CSIRO Chair of Excellence Medal, The Commonwealth Scientific and Industrial Research Organisation (CSIRO); Australia's national science agency
- 2014 The Cinda Helke Award for Excellence in Graduate Student Admissions
- 2016 The James J. Leonard Award for Excellence in Translational/Clinical Research
- 2019 The 2019 Federal Laboratory Consortium (FLC) Award for Excellence in Technology Transfer
- 2019 USU Outstanding Biomedical Graduate Educator Award

C. Contributions to Science

My Ph.D. thesis revealed that *S. pneumoniae* elaborated surface receptors for human plasmin on Group A *Streptococci* during a rotation of fibrinolytic enzyme, plasmin, towards the zymogen precursor plasminogen.

other serine proteases. This bacterium binds to fibrin and remains enzymatically active throughout its ability to hydrolyze a fibrin cross-linker. This bacterium is produced as a natural byproduct of the bacterium producing an active enzyme that destroys the extracellular matrix environment, first and foremost, the "flesh-eating streptococci".

- a. Lottenberg R, Broder CC, Rowe MDP (1987). Identification of a Specific Receptor for Plasmin on a Group A Streptococcus. *Infection and Immunity* 55(6): 1511-1516.
- b. Broder CC, Lottenberg R, Rowe MDP (1988). Mapping of a Specific Receptor for Plasmin Recognized by This Unique Group A Streptococcus. *Infection and Immunity* 48(1): 260-265.
- c. Broder CC, Lottenberg R, von Mennd GO, Johns R (1988). A Specific Plasmin Receptor Produced by the Same Microorganism. *J. Biol. Chem.* 263:4922-28.
- d. Lottenberg R, Broder CC, Rowe MDP, Keis S, Schroeder DL, Curtiss III R (1988). Sequence Analysis, and Expression in *Escherichia coli* of a Streptococcal Plasmin Receptor. *J. Bacteriology* 174:5264-5270.

2. My independent postdoctoral fellowship focused on the early stages of HIV-1 infection. I generated the first panel of T-cell and macrophage-tropic HIV-1 envelope glycoprotein (Env) encoding recombinant vaccinia virus vectors and used these tools to study the cellular tropism of HIV-1. I also developed the first soluble glycoprotein and analyzed the immunogenic structure in terms of its presentation conformational and virus-neutralizing epitopes through the development and characterization of more than 100 murine monoclonal antibodies.

- a. Broder CC, Diamond D, Berger EA (1986). The Block to HIV-1 Envelope Glycoprotein-Mediated Membrane Fusion in Animal Cells Expressing Human CD4 Cell Surface Receptor by a Human Cell Component. *J. Virol.* 57:193-198.
- b. Nussbaum O, Broder CC, Berger EA (1988). HIV-1 Envelope Glycoprotein/CD4 Mediated Cell Fusion: A Novel Receptor. *Infection and Immunity* 48(1): 260-265.
- c. Broder CC, Earl PL, Lottenberg R, Doms RW (1994). Antigenic Implications of HIV-1 Envelope Glycoprotein Quaternary Structure: Oligomer Specific and Sensitive. *PNAS* 91:11600-11605.
- d. Broder CC, Berger EA (1995). Fusogenic Selectivity of the Envelope Glycoprotein Determinant of HIV-1 Tropism for CD4+ T Cell Lines vs. Macrophages. *PNAS* 92:1000-1005.

3. My independent postdoctoral fellowship focused on the cellular and viral membrane fusion tropism of HIV-1 and the development of a sensitive and specific assay for cell-cell membrane fusion. I hypothesized existed, and this was the first time that a specific receptor for macrophage-tropic Envs (CCR5) was identified. The identification of CCR5 as a HIV-1 receptor led to the development of a sensitive and specific assay for cell-cell membrane fusion. This assay was used to study the cellular and viral membrane fusion tropism of HIV-1 and the development of therapeutic strategies.

- a. Feng Y, Broder CC, Kennedy PE, Berger EA (1996). HIV-1 Entry Coceptor: Functional CDNA Cloning of a Seven-Transmembrane G Protein-Coupled Receptor. *Science* 272:1955-1958.
- b. Alkhalil O, Sambasivan S, Broder CC, Feng Y, Kennedy PE, Murphy PM, Berger EA (1996). CC CKR5: a Novel HIV-1 Receptor. *PNAS* 93:11600-11605.

- c. Rucker J, Samson M, Doranz BJ, Libert F, Berson A, Kim Y, Collman RG, Broder CC, Doms R (2007). *Structural Basis of HIV-1 Coreceptor Specificity*. *Cell* 127:119-131
- d. Edinger AL, Amédecé A, Miller K, Long E, Foy J, Ehrlich M, Stransky M, Samson M, Cuzin, Clements JE, Murphy-Korwini M, Peiper SC, Parmentier M, Broder CC (2007). Differential utilization of CXCR5 by macrophage and T cell-tropic simian immunodeficiency virus strains. *PNAS USA*, 104:1005-1010

My most recent work on HIV-1 entry led to further independent investigations characterizing the role of the HIV-1 coreceptors in the viral entry process. These studies revealed the interplay between the HIV-1 entry receptors, spanning junctions of the coreceptors involved in HIV-1 Env interaction, and also revealed possible avenues of how an HIV-1 Env might engage an alternatively used CXCR4 and CXCR5 coreceptor. I have also been collaborating with a group of researchers to follow-up studies exploring the utility of soluble oligomeric HIV-1 envelope glycoprotein subunit vaccination immunogens (gp140) which initiated a neutralizing antibody response with the unusual R2 HIV-1 Env isolate and led to the development of a novel HIV-1 vaccine.

- a. Chabot DJ, Zhang PF, Quinlan CV, Broder CC (2000). Mutagenesis of CXCR4 Reveals Cryptic Coreceptor Activity for HIV-1 Isolates. *J Virol* 74:1335-1344
- b. Xiao X, Wu L, Sun Y, Chen H, Wang Y, Eggen S, Chen H, Chen L, Broder CC, Gammie A (2007). *Structural Basis of Coreceptor Surface Association between CD4 and CXCR4*. *PNAS USA* 104:7500-7505
- c. Chabot DJ, Chen H, Ding W, Broder CC (2006). *N-linked Glycosylation in CXCR4 Masks Coreceptor Function for CXCR5-Directed HIV-1 Infection*. *J Virol* 80:1411-1420
- d. Zhang PF, Chen H, Dong M, Broder CC, Poulos J, Broder CC, Quinlan CV, Broder CC, N, Voss G, Broder CC, Quinlan CV (2007). Extensively cross-reactive anti-HIV-1 neutralizing antibodies induced by gp140 immunization. *PNAS USA* 104(24):10193-8

5. My most recent research has been on emerging viruses that impact human and domestic livestock populations. I have been studying Hendra virus (HeV) and Nipah virus (NiV) in Australia and the Philippines. My lab was the first to identify HeV in Australia. I obtained the first NIAID funded project providing monetary support to an overseas laboratory (2003-2007) for 5 years, covering the development and testing of fusion inhibitors, subunit vaccine and neutralizing human monoclonal antibody (mAb) and supported the development of the first and only HeV and NiV vaccine in Australia and the development of the first non-human primate model for the *in vivo* efficacy of the Hendra/Nipah vaccine (HeV-sG) and an anti-HeV/NiV G-specific neutralizing human mAb. One human mAb (m1C5) was used by compassionate emergency protocols in 10 people in Australia and one individual in the United States (2004-2005). In 2016, the henipavirus subunit vaccine, HeV-sG, was launched; called Equivac® HeV (Zoetis, Inc.) and is the first vaccine against a lethal disease. My lab also identified the structure of the henipavirus entry receptors (ephrin-B2/B3) and produced soluble versions of the G and F glycoproteins facilitating their structural solutions.

- a. Bossart KN, Zhu Z, Middleton D, Klippel J, Crameri G, Bingham J, McEachern JA, Green D, Hancock TJ, Chan Y-P, Hickey AC, Dimitrov DS, Wang L-P, Broder CC (2009). A neutralizing human monoclonal antibody protects against lethal disease in a new model of acute Nipah virus infection. *J Virol* 83:1052-1060
- b. Bossart KN, Geisbert TW, Broder CC, Chan Y-P, Hickey AC, Dimitrov DS, Broder CC, Scott MG, Wang Y, Dimitrov AS, Callison J, Chan Y-P, Hickey AC, Dimitrov DS, Broder CC (2016). Structure of the Henipavirus G Glycoprotein. *J Virol* 90:1052-1060

- 2005-06 Medical Institute student intern, College Park, MD.
- 2005-06 National Institute of Neurological Disorders and Stroke, College Park, MD.
- 2005-06 Maryland College Park, MD.
- 2007-08 Undergraduate research assistant, Biology Department, Uniformed Services University, Bethesda, MD.
- 2007-08 University of Maryland, College Park, MD.
- 2008-09 Research assistant, Department of Pharmacology, Uniformed Services University, Bethesda, MD.
- 2010 Research assistant, Department of Microbiology, Uniformed Services University, Bethesda, MD.
- 2010-16 Graduate research student, Department of Microbiology, Uniformed Services University, Bethesda, MD.
- 2016-17 Postdoctoral Fellow, Henry M. Jackson Foundation, Department of Microbiology, Uniformed Services University, Bethesda, MD.
- 2017-18 Scientist II, Molecular Biology Department, Uniformed Services University, Bethesda, MD.
- 2019 - Research Assistant Professor, Uniformed Services University, Department of Microbiology and Immunology, Uniformed Services University, Bethesda, MD.

Other Experience and Professional Membership

- 2009 Mentor, A&P Student mentoring, Bethesda City, Grade High School, Bethesda, MD.
- 2010-10 Mentor, high school, undergraduate, and graduate students, Uniformed Services University, Bethesda, MD.
- 2014 Participant, AAAS/American Society of Microbiology
- 2014-15 Volunteer, AAAS/Society Scientists and Engineers STEM Volunteer Program
- 2014-17 Member, American Society of Tropical Medicine and Hygiene
- 2014-19 Member, American Society of Microbiology
- 2015-16 Member, USU Global Health Interest Group

Honors

- 2005-07 Semester Honors
- 2006 College Park Life Sciences Scholars Program
- 2008 High Honors, Biology Departmental Honors Program
- 2015 USU Research Days Graduate Student Award
- 2015 NSF East Asia and Pacific International Research Student Award
- 2015-16 Val G. Hemming Fellowship, Henry M. Jackson Foundation

C. Career Summary

1. **Virus-host interactions.** My Ph.D. thesis research was focused on virus-host interactions, understanding bats as hosts of zoonotic viruses and Australian bat lyssavirus (ABLV) cellular entry, exploring the antiviral mechanisms that enable cellular persistence of viruses in bats, particularly, autophagy. Findings revealed that the autophagy pathway is induced upon infection with Australian bat lyssavirus (ABLV), a rabies virus-related virus carried by Australian bats. Pharmacological and genetic studies of the autophagy pathway in the ABLV-infected cells indicated that autophagy functions as an antiviral defense. The study also demonstrated that bat-derived cell lines have elevated levels of basal autophagy, which may contribute to the ability of bats to act as hosts to these viruses. An additional finding from these studies was that activation of autophagy in mammalian cells confers resistance to ABLV infection.

Wellcome Trust: Tropical Medicine Interest Group, Member

Member

Gates Foundation: Founding Board, Global Alliance to Improve Nutrition, Grand Challenges in Global Health, Scientific Advisory Board, International Scientific Advisory Committee, Member

One Health Consortium, ISORS

Consortium of Universities for Global Health

National Center for Genetic Engineering and Biotechnology, National Science and Technology Development

Institute for Healthcare Improvement, Scientific Advisor

Council on Health Research and Statistics, Member

Nevin Scrimshaw International Nutrition Foundation, Member

American Federation for Clinical Research: Member

American Society for Microbiology: Member

American Association for the Advancement of Science: Member

American Society for Clinical Investigation: Member

Association of American Physicians: Member

New York Academy of Sciences: Member

Honors

1972 - American Board of Internal Medicine, Diplomate, Internal Medicine

1973 -76 Career Scientist Award, Health Research Council

1974 -79 Research Career Development Award, NIAID

1981 Oswald Avery Award, Infectious Diseases Society of America

1991 Health Officer, Visiting Professor, University

1997 Maxwell Finland Lectureship, Infectious Disease Society of America

2000 Edward R. Basky Award, Physicians Forum/Physicians for Social Responsibility

2002 Alexander Fleming Award, Infectious Diseases Society of America

2002 National Academy of Medicine, Elected Member

2005 Robert Rousement Award, American Academy of Microbiology

2013 Distinguished Leadership Award, American Society for Microbiology

Contributions to Science

1. I rediscovered Shiga toxin during my fellowship, while participating in a training program at the Institute of Nutrition for Central America and Panama in Panama. This work has focused on Shiga toxin's ability to invade epithelial cells, the previously described damage of the bowel and body inflammatory exudates due to dysentery in a rabbit model. I proved the two toxin activities were due to the same protein. My work on Shiga toxin (Stx), sequenced the binding subunit, identified its mammalian cell receptor, described its translocation to the cell cytoplasm via receptor. This work paved the way to understand the pathogenesis of *E. coli* O157 and other serotypes associated with hemorrhagic colitis and hemolytic uremic syndrome. My research was essential for a rapid commercial diagnostic test for all Stx-producing bacteria. I was principal investigator of all of these studies.

a. Keusch GT, Grady GF, Mata JM, McIver JM (1972) The pathogenesis of *Citrobacter* Enterocolitis

- 1982 -94 Associate Professor of Immunology, Boston University School of Medicine
- 1994 - Professor and Chair, Department of Immunology, Boston University School of Medicine
- 2007 -14 Associate Director, National Emerging Infectious Diseases Laboratory, Boston University
- 2009 -14 Associate Professor of Research, Boston University Medical Campus
- 2014 - Director, National Emerging Infectious Diseases Laboratory, Boston University

Other Experience and Professional Memberships

- 1988 -92 Immunobiology Study Section, NIH
- 1996 -00 Immunobiology Study Section, NIH
- 2004 Immunobiology Study Section, NIH
- 1978 - American Association of Immunologists
- 1991 - American Society for Microbiology
- 1992 -93 American Society for Microbiology
- 1997 -98 Chair, Cell Biology
- 2000 Chair, Cell Biology and Immunology Predoctoral Committee, HHMI
- 2001 -03 Member, Research Training Fellowships for Medical Students Committee, HHMI
- 2002 Member, Research Training Fellowships for Medical Students Committee, HHMI
- 2002 Advisory Panel, NIH
- 2005 -06 Chair, "Med Interscience Initiatives" Panel, HHMI
- 2007 Member, NIH Review Panels on "B Cell Immunology and Effective HIV-1 Vaccines"
- 2009, 10 Member, NIH Review Panels on "Basic HIV Discovery Research"
- 2011 -13 Member, ad hoc Review Panel, "Immune Mechanisms of Virus Control", HHMI
- 2011 -14 SmithGroup JJR Science & Technology Advisory Board
- 2016, 17 Reviewer, National Research Foundation, Competitive Research Program, Singapore
- Ongoing: Security Risk Assessment (SRA) Select Agents and Toxins (SAT)
- Ongoing: BSL-4 suit-trained and certified, Boston University

Honors

- 1979 - Fellow of the American Society for Microbiology
- 2015 - Fellow of the American Association of Immunologists

C. Career

1. **Innate role of B lymphocytes**
secreted IgM antibodies had unique functions in concentrating pathogens and antigen into secondary lymphoid organs, and prevented dissemination into vital organs. We sought to determine the cells responsible for these activities, and to understand the consequences for the immune system. We demonstrated that IgM immune complexes became co-packaged with B cells, and were transported these complexes to the spleen. We discovered that B cells have an innate role for this subset of lymphocytes in the early steps of initiation of primary immune responses. A role for orchestrated transport of antigen and immune complexes to the spleen is now widely accepted as early events in immune responses.
 - a. Ferguson AR, Corley RB (2008). Accumulation of marginal zone B cells and accelerated loss of follicular dendritic cells in TNF- α deficient mice. *BMC Immunology* 6:8
 - b. Ferguson AR, Youd ME, Corley RB (2007). Marginal zone B cells transport immune complexes of follicular dendritic cells to the spleen ("immune complex mouth").
 - c. Youd ME, Ferguson AR, Corley RB (2002). Synergistic roles of IgM immune complexes and follicular localization. *Eur. J. Immunol.* 32:233-237.

The fact that a subset of IgM antibodies in immune responses... and others have indicated that IgM antibodies were not always secreted as pentameric molecules... chain, but... evidence existed to indicate... functions with pentameric IgM, or if they had unique functions... monomers, had discrete activities... could be deleterious... autoimmune diseases... monomers did not fix complement... ability to function in antigen trapping, and could also accelerate disease manifestations... autoimmune prone mice. These data supported the important role for strict quality control standards in the assembly and secretion of IgM antibodies for maintenance of proper homeostasis in the immune system.

- a. Youd ME, Luus L, Corley RB (1997). IgM hexamers in autoimmune disease: one fas-deficient mice. *J. Autoimmunity* 10: 333-343.
- b. Hughey CT, Brewer JW, Corley RB (1997). Normal and autoimmune B cells: Implications for the physiological role of hexameric IgM. *J. Immunol.* 158: 4091-4097.
- c. Brewer JW, Corley RB (1997). Late secretory intracellular IgM polymerization: activity of secretory IgM. *Mol. Immunol.* 34: 221-228.
- d. Brewer JW, Randall TD, Parkhouse RML, Corley RB (1994). IgM hexamers. *Immunol. Today* 15: 165-168.

3. Quality control in modulating the assembly and secretion of IgM

controlling the assembly and secretion of IgM... whether the addition... was responsible for catalyzing assembly of IgM into... controversial. We demonstrated, however, that IgM assembly is regulated in the endoplasmic reticulum by a process involving thiol regulation. Further, J chain plays a role in mediated IgM assembly and its addition is a terminally late step in the production of polymeric IgM. To complete these studies, we made use of various biochemical assays including pulse chase experiments. We also cloned and expressed J chain to demonstrate its role in modulating polymer assembly, and this work remains the definitive description of IgM assembly.

- a. Reddy PS, Corley RB (1999). The role of J chain in the assembly and secretion of secretory IgM. *Immunol. Rev.* 20: 582-588.
- b. Brewer JW, Corley RB (1997). The regulation of protein synthesis in the endoplasmic reticulum. *Immunol. Rev.* 160: 107-118.
- c. Brewer JW, Randall TD, Parkhouse RML, Corley RB (1994). Mechanisms of and substrate for J chain mediated IgM polymer assembly. *J. Biol. Chem.* 269: 17336-17340.
- d. Randall TD, Brewer JW, Corley RB (1992). Direct evidence that J chain regulates the polymeric structure of IgM in antibody secreting B cells. *J. Biol. Chem.* 267: 10062-10067.

4. Defining mouse mammary tumor virus as an endogenous

of B lymphocytes in the MMTV life cycle. Prior to these studies there was evidence for the existence of that endogenous superantigens which played important roles in shaping the T cell repertoire in mice, but the identity of these superantigens was unknown. In the course of our studies on the MMTV life cycle, we identified mouse mammary tumor virus, MMTV, as a superantigen that mediated the activation of B cells, and had linked this to superantigen mediated activation of B cells in the MMTV life cycle.

- a. Sharma S, King LR, Corley RB (1998). Identification of endogenous mouse mammary tumor proviral env transcripts following B cell stimulation. *J. Immunol.* 161: 2510-2518.

1;110(40):16157-62.

- 4. Menachery VD, Yount RL, Sims A, Agriyilatharan S, Gralinski LE, Plante IA, Graham RM, Seay Royal S, Pickles RJ, Randell SH, Lera-Alecio A (2017). WIV1-CoV poised for human emergence. **PNAS** 15:113(11):3048-53.

B. Positions and Employment

Positions and Employment

- 1993 American Society for Microbiology
- 1994 Albert Einstein College of Medicine Summer Student Award
- 1996-01 Graduate Student, Laboratory of Mark Denison, Vanderbilt University, Nashville, TN
- 1999 Dissertation Enhancement Award, Vanderbilt University
- 2001-02 Postdoctoral Fellow, Duke University
- 2002-04 Infectious Disease Pathogenesis Training Grant Fellow
- 2002-05 Postdoctoral Fellow, University of North Carolina at Chapel Hill
- 2005-17 Research Assistant Professor, Department of Food Bioprocess Technology
- 2017- Present

C. Contributions

1. **In vitro model of human airway epithelium** - The primary human airway epithelium is a complex, multi-layered structure that is difficult to culture in vitro. We have developed a primary human conducting airway can be cultured at an air liquid interface and following maturation recapitulate the morphology of the airway epithelium. These cultures provide a unique in vitro model and for studying human coronavirus HKU1 provide the only in vitro model for studying this virus

- a. **Sims A, Pvrk K, Dijkman R, Jebbink M, Lond C, Demind D, De Boer E, Vabret A, Bañic R, van der Hoek L, Pickles R** (2010). Culturing the unculturable: human coronavirus HKU1 infects, replicates, and produces progeny virions in human airway epithelial cell cultures. **J Virol** 84(27):1200-03.
- b. **Sims A, Bañic RS, Yount R, Dijkman R, Demind D, Pvrk K, Pickles R** (2009). SARS-CoV infection of human ciliated airway epithelium: the role of the ciliated cell in viral spread in the conducting airways of the lung. **J Virol** 79(24):15524-31.
- c. **Uygun Y, Dora W, Milovanovic A, Galda A, Qi Y, Zhu Q, Marasco W, Bañic R, Sims A, van der Hoek L, Sai J*** (2015). HCoV-NL63 RNA-proteinase C sequestered acidic organelles as a determinant and employs HE protein as a receptor-destroying enzyme. **Virology** 484(1-2):102-19.
*indicates co-senior author

2. **Development of coronavirus infectious clones**. The development of coronavirus infectious clones has drastically increased the understanding of how specific genes or open reading frames direct replication and pathogenesis, as well as identifying sets of mutations that can make coronavirus genomes more proof live.

- a. Thornbrough JM, Jha BK, Yount B, Goldstein M, Weiss SR (2016). Middle East Respiratory Syndrome Coronavirus NS4b Protein Inhibits Host RNase I Activation. **MBio** 29:7(2)
- b. Menachery VD, Gralinski LE, Mitchell DM, Plante IA, Yount RL, Graham RM, Seay Royal S, Pickles RJ, Randell SH, Lera-Alecio A, ET, Stratton KG, Cockrell AS, DeBorja R, Sims A, Waters KM, Bañic RS (2017). Middle East Respiratory Syndrome Coronavirus Nonstructural Protein 16 is Necessary for Interferon Persistence and Viral Pathogenesis. **mSphere** 2(6):e00346-17.

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